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## IMPROVED VESICLES

All documents cited herein are incorporated by reference in their entirety.

### TECHNICAL FIELD

This invention is in the field of vesicle preparation for immunisation purposes.

### 5 BACKGROUND ART

One of the various approaches to immunising against *N.meningitidis* infection is to use outer membrane vesicles (OMVs). An efficacious OMV vaccine against serogroup B has been produced by the Norwegian National Institute of Public Health [e.g. ref. 1] but, although this vaccine is safe and prevents NmB disease, its efficacy is limited to the strain used to make the vaccine.

- 10 The 'RIVM' vaccine is based on vesicles containing six different PorA subtypes and has been shown to be immunogenic in children in phase II clinical trials [2].

Reference 3 discloses a vaccine against different pathogenic serotypes of serogroup B meningococcus based on OMVs which retain a protein complex of 65-kDa. Reference 4 discloses a vaccine comprising OMVs from genetically-engineered meningococcal strains, with the OMVs  
15 comprising: at least one Class 1 outer-membrane protein (OMP) but not comprising a Class 2/3 OMP. Reference 5 discloses OMVs comprising OMPs which have mutations in their surface loops and OMVs comprising derivatives of meningococcal lipopolysaccharide (LPS).

- Reference 6 discloses compositions comprising OMVs supplemented with transferrin binding proteins (e.g. TbpA and TbpB) and/or Cu,Zn-superoxide dismutase. Reference 7 discloses  
20 compositions comprising OMVs supplemented by various proteins. Reference 8 discloses preparations of membrane vesicles obtained from *N.meningitidis* with a modified *fur* gene.

- As well as serogroup B *N.meningitidis*, vesicles have been prepared for other bacteria. Reference 9 discloses a process for preparing OMV-based vaccines for serogroup A meningococcus. References  
10 and 11 disclose vesicles from *N.gonorrhoeae*. Reference 12 discloses vesicle preparations from  
25 *N.lactamica*. Vesicles have also been prepared from *Moraxella catarrhalis* [13,14], *Shigella flexneri* [15,16], *Pseudomonas aeruginosa* [15,16], *Porphyromonas gingivalis* [17], *Treponema pallidum* [18], *Haemophilus influenzae* [19 & 20] and *Helicobacter pylori* [21].

- One drawback with bacterial vesicle preparations is that important protective antigens are not present. To retain antigens such as NspA in OMV preparations, reference 20 teaches that *nspA*  
30 expression should be up-regulated with concomitant *porA* and *cps* knockout. It is an object of the invention to provide further and improved vesicle preparations, together with processes for their manufacture. In particular, it is an object of the invention to provide vesicles which retain important bacterial immunogenic components from *N.meningitidis*.

## DISCLOSURE OF THE INVENTION

Prior art methods of meningococcal OMV preparation involve the use of detergent during disruption of the bacterial membrane [e.g. see ref. 22, where a deoxycholate detergent is used]. The invention is based on the surprising discovery that membrane disruption substantially in the absence of detergent results in OMVs which retain important bacterial immunogenic components, particularly strain-to-strain conserved components such as (i) the protective NspA surface protein, (ii) protein '287' and (iii) protein '741'.

Therefore the invention provides a process for the manufacture of an outer membrane vesicle preparation from a bacterium, wherein the bacterial membrane is disrupted substantially in the absence of detergent. OMV preparations obtainable by processes of the invention are also provided.

For obtaining NspA<sup>+</sup> vesicles, the process of the invention is much simpler than performing multiple genetic manipulations as described in reference 20.

The process of the invention will typically involve the following basic steps: (a) treating bacterial cells in the substantial absence of detergent; (b) centrifuging the composition from step (a) to separate the outer membrane vesicles from treated cells and cell debris, and collecting the supernatant; (c) performing a high speed centrifugation of the supernatant from step (b) and collecting the outer membrane vesicles in a pellet; (d) re-dispersing the pellet from step (c) in a buffer; (e) performing a second high speed centrifugation in accordance with step (c), collecting the outer membrane vesicles in a pellet; (f) re-dispersing the pellet from step (e) in an aqueous medium.

The process may also comprise the following steps: (g) performing sterile filtration through at least two filters of decreasing pore size of the re-dispersed composition from step (f); and (h) optionally including the composition from step (g) in a pharmaceutically acceptable carrier and/or adjuvant composition.

Step (a) gives rise to vesicles of the bacterial outer membrane, and the vesicles generally comprise outer membrane components in substantially their native form. Advantageously, strain-to-strain membrane components NspA, '287' and '741' are preserved.

Step (b) will typically involve centrifugation at around 5000 – 10000 *g* for up to 1 hour. Steps (c) and (e) will typically involve centrifugation at around 35000 – 100000 *g* for up to 2 hours. Centrifugation steps are preferably performed at between 2°C and 8°C.

Any suitable buffer can be used in step (d) e.g. Tris buffer, phosphate buffer, histidine buffer *etc.* Step (f) may also involve the use of a buffer, which may be the same buffer as used in step (d), or may simply involve the use of water (e.g. water for injection).

Step (g) preferably ends with a filter of pore-size of about 0.2µm.

The invention also provides a *N.meningitidis* vesicle composition, characterised in that the vesicles include (i) NspA protein, (ii) '287' protein and (iii) '741' protein.

### *The bacterium*

The bacterium from which OMVs are prepared may be Gram-positive, but it is preferably Gram-negative. The bacterium may be from genus *Moraxella*, *Shigella*, *Pseudomonas*, *Treponema*, *Porphyromonas* or *Helicobacter* (see above for preferred species) but is preferably from the

5 *Neisseria* genus. Preferred *Neisseria* species are *N.meningitidis* and *N.gonorrhoeae*. Within *N.meningitidis*, any of serogroups A, C, W135 and Y may be used, but it is preferred to prepare vesicles from serogroup B. Preferred strains within serogroup B are MC58, 2996, H4476 and 394/98.

To reduce pyrogenic activity, it is preferred that the bacterium should have low endotoxin (LPS) levels. Suitable mutant bacteria are known *e.g.* mutant *Neisseria* [23] and mutant *Helicobacter* [24].

10 Processes for preparing LPS-depleted outer membranes from Gram-negative bacteria are disclosed in reference 25

The bacterium may be a wild-type bacterium,<sup>1</sup> or it may be a recombinant bacterium. Preferred recombinant bacteria over-express (relative to the corresponding wild-type strain) immunogens such as NspA, 287, 741, TbpA, TbpB, superoxide dismutase [6], *etc.* The bacterium may express more

15 than one PorA class I outer membrane protein *e.g.* 2, 3, 4, 5 or 6 of PorA subtypes: P1.7,16; P1.5,2; P1.19,15; P1.5c,10; P1.12,13; and P1.7h,4 [*e.g.* refs. 26, 27].

The process of the invention will typically involve an initial step of culturing the bacteria, optionally followed by a step of concentrating the cultivated cells.

### *Membrane disruption*

20 Membrane disruption for vesicle formation is performed substantially in the absence of detergent.

In particular, membrane disruption may be performed substantially in the absence of a deoxycholate detergent, with other detergents optionally being present.

Membrane disruption may be performed substantially in the absence of ionic detergent, with non-ionic detergent optionally being present. Alternatively, it may be performed substantially in the

25 absence of non-ionic detergent, with ionic detergent optionally being present. In some embodiments, neither ionic nor non-ionic detergent is present.

Steps after membrane disruption and vesicle formation may involve the use of detergent. Thus a process wherein membrane disruption occurs in the absence of detergent, but in which detergent is later added to the prepared vesicles, is encompassed within the invention.

30 The term "substantially in the absence" means that the detergent in question is present at a concentration of no more than 0.05% (*e.g.*  $\leq 0.025\%$ ,  $\leq 0.015\%$ ,  $\leq 0.010\%$ ,  $\leq 0.005\%$ ,  $\leq 0.002\%$ ,  $\leq 0.001\%$  or even 0%) during membrane disruption. Thus processes where trace amounts of detergent are present during vesicle preparation are not excluded.

Membrane disruption in the absence of detergent may be performed on intact bacteria using physical techniques *e.g.* sonication, homogenisation, microfluidisation, cavitation, osmotic shock, grinding, French press, blending *etc.*

### *The vesicles*

- 5 The processes of the invention produce outer membrane vesicles. OMVs are prepared from the outer membrane of cultured bacteria. They may be obtained from bacteria grown in broth or in solid medium culture, preferably by separating the bacterial cells from the culture medium (*e.g.* by filtration or by a low-speed centrifugation to pellet the cells), lysing the cells (without detergent), and separating an outer membrane fraction from cytoplasmic molecules (*e.g.* by filtration, by differential precipitation or aggregation of outer membranes and/or OMVs, by affinity separation methods using
- 10 ligands that specifically recognize outer membrane molecules, or by a high-speed centrifugation that pellets outer membranes and/or OMVs).

- OMVs can be distinguished from microvesicles (MVs [28]) and 'native OMVs' ('NOMVs' [63]), which are naturally-occurring membrane vesicles which form spontaneously during bacterial growth and are released into culture medium. MVs can be obtained by culturing *Neisseria* in broth culture medium, separating whole cells from the broth culture medium (*e.g.* by filtration or by low-speed centrifugation to pellet only the cells and not the smaller blebs) and then collecting the MVs that are present in the cell-depleted medium (*e.g.* by filtration, by differential precipitation or aggregation of MVs, by high-speed centrifugation to pellet the MVs). Strains for use in production of MVs can
- 15 generally be selected on the basis of the amount of MVs produced in culture. References 29 and 30 describe *Neisseria* with high MV production.

### *Retained bacterial immunogenic components*

- The substantial absence of detergent in processes of the invention results in vesicle preparations which retain immunogenic components of the bacterial surface which, using detergent-based prior art
- 25 methods, would otherwise be lost or decreased. In *N.meningitidis*, three immunogens which are advantageously retained using the invention include, but are not limited to: (1) NspA; (2) protein '741'; and (3) protein '287'.

- NspA (Neisserial surface protein A) is disclosed in references refs. 31 to 37 and as SEQ IDs 4008-4033 of reference 38. It is a candidate vaccine for the prevention of meningococcal disease. It is
- 30 highly conserved between strains. Despite initial hope, however, it is now believed that NspA will not be an adequate protective antigen on its own and will need to be administered with additional antigens [*e.g.* ref. 36, and example 11 of ref. 38]. NspA has been found to be removed by prior art detergent-based preparation methods. According to the present invention, however, NspA can be retained in vesicles. Such NspA<sup>+</sup> vesicles are advantageous because a combination of two known
- 35 potent immunogens (*i.e.* vesicles + NspA) is prepared in a single process, with each immunogen enhancing the efficacy of the other.



Protein '741' is disclosed as 'NMB1870' in reference 39 (GenBank: AAF42204, GI:7227128). It is also disclosed in references 40 and 41. It elicits strong bactericidal antibodies. It has been found that protein '741' is partially removed in vesicles prepared by prior art detergent-based methods. According to the present invention, however, '741' can be retained in vesicles. Such 741<sup>+</sup> vesicles are advantageous because a combination of two known potent immunogens (*i.e.* vesicles + 741) is prepared in a single process, with each immunogen enhancing the efficacy of the other.

Protein '287' is disclosed as 'NMB2132' in reference 39 (GenBank: AAF42440, GI:7227388). It is also disclosed in references 40 and 42. It elicits strong bactericidal antibodies. Protein '287' is typically not present in vesicles prepared by prior art detergent-based methods and, to overcome its removal, it has previously been proposed that OMV preparations might be supplemented with 287 [43]. According to the present invention, however, '287' can be retained in vesicles. Such 287<sup>+</sup> vesicles are advantageous because a combination of two known potent immunogens (*i.e.* vesicles + 287) is prepared in a single process, with each immunogen enhancing the efficacy of the other.

Preferred NspA (a) has at least *a*% sequence identity to amino acid sequence GI:1518522 and/or (b) comprises a fragment of at least *x* amino acids from amino acid sequence GI:1518522. Preferred '741' (a) has at least *b*% sequence identity to amino acid sequence GI:7227128 and/or (b) comprises a fragment of at least *x* amino acids from amino acid sequence GI:7227128. Preferred '287' (a) has at least *c*% sequence identity to amino acid sequence GI:7227388 and/or (b) comprises a fragment of at least *x* amino acids from amino acid sequence GI:7227388. The values of *a*, *b* and *c* are independent from each other, but each value is at least 70 (*e.g.* 75, 80, 85, 90, 95, 96, 97, 98, 99, 99.5 or 100). The values of *x*, *y* and *z* are independent from each other, but each value is at least 8 (*e.g.* 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 *etc.*). Fragments preferably comprise epitopes.

Preferred NspA, 287 and 741 proteins substantially retain the ability of the wild-type proteins (as found in intact bacteria) to elicit bactericidal antibodies in patients.

#### *Immunogenic pharmaceutical compositions*

The process of the invention provides a vesicle preparation. For administration to a patient, the vesicles are preferably formulated as immunogenic compositions, and more preferably as compositions suitable for use as a vaccine in humans (*e.g.* children or adults). Vaccines of the invention may either be prophylactic (*i.e.* to prevent infection) or therapeutic (*i.e.* to treat disease after infection), but will typically be prophylactic.

The composition of the invention is preferably sterile.

The composition of the invention is preferably pyrogen-free.

The composition of the invention generally has a pH of between 6.0 and 7.0, more preferably to between 6.3 and 6.9 *e.g.* 6.6±0.2. The composition is preferably buffered at this pH.

Other components suitable for human administration are disclosed in reference 44.

The composition will generally comprise an adjuvant. Preferred adjuvants to enhance effectiveness of the composition include, but are not limited to: (A) MF59 (5% Squalene, 0.5% Tween 80, and 0.5% Span 85, formulated into submicron particles using a microfluidizer) [see Chapter 10 of ref. 45; see also ref. 46]; (B) microparticles (*i.e.* a particle of ~100nm to ~150µm in diameter, more preferably ~200nm to ~30µm in diameter, and most preferably ~500nm to ~10µm in diameter) formed from materials that are biodegradable and non-toxic (*e.g.* a poly( $\alpha$ -hydroxy acid), a polyhydroxybutyric acid, a polyorthoester, a polyanhydride, a polycaprolactone *etc.*), with poly(lactide-co-glycolide) being preferred, optionally being charged surface (*e.g.* by adding a cationic, anionic, or nonionic detergent such as SDS (negative) or CTAB (positive) [*e.g.* refs. 47 & 48]); (C) liposomes [see Chapters 13 and 14 of ref. 45]; (D) ISCOMs [see Chapter 23 of ref. 45], which may be devoid of additional detergent [49]; (E) SAF, containing 10% Squalene, 0.4% Tween 80, 5% pluronic-block polymer L121, and thr-MDP, either microfluidized into a submicron emulsion or vortexed to generate a larger particle size emulsion [see Chapter 12 of ref. 45]; (F) Ribi<sup>TM</sup> adjuvant system (RAS), (Ribi Immunochem) containing 2% Squalene, 0.2% Tween 80, and one or more bacterial cell wall components from the group consisting of monophosphorylipid A (MPL), trehalose dimycolate (TDM), and cell wall skeleton (CWS), preferably MPL + CWS (Detox<sup>TM</sup>); (G) saponin adjuvants, such as QuilA or QS21 [see Chapter 22 of ref. 45], also known as Stimulon<sup>TM</sup>; (H) chitosan [*e.g.* 50]; (I) complete Freund's adjuvant (CFA) and incomplete Freund's adjuvant (IFA); (J) cytokines, such as interleukins (*e.g.* IL-1, IL-2, IL-4, IL-5, IL-6, IL-7, IL-12, *etc.*), interferons (*e.g.* interferon- $\gamma$ ), macrophage colony stimulating factor, tumor necrosis factor, *etc.* [see Chapters 27 & 28 of ref. 45]; (K) a saponin (*e.g.* QS21) + 3dMPL + IL-12 (optionally + a sterol) [51]; (L) monophosphoryl lipid A (MPL) or 3-O-deacylated MPL (3dMPL) [*e.g.* chapter 21 of ref. 45]; (M) combinations of 3dMPL with, for example, QS21 and/or oil-in-water emulsions [52]; (N) oligonucleotides comprising CpG motifs [53] *i.e.* containing at least one CG dinucleotide, with 5-methylcytosine optionally being used in place of cytosine; (O) a polyoxyethylene ether or a polyoxyethylene ester [54]; (P) a polyoxyethylene sorbitan ester surfactant in combination with an octoxynol [55] or a polyoxyethylene alkyl ether or ester surfactant in combination with at least one additional non-ionic surfactant such as an octoxynol [56]; (Q) an immunostimulatory oligonucleotide (*e.g.* a CpG oligonucleotide) and a saponin [57]; (R) an immunostimulant and a particle of metal salt [58]; (S) a saponin and an oil-in-water emulsion [59]; (T) *E.coli* heat-labile enterotoxin ("LT"), or detoxified mutants thereof, such as the K63 or R72 mutants [*e.g.* Chapter 5 of ref. 60]; (U) cholera toxin ("CT"), or detoxified mutants thereof [*e.g.* Chapter 5 of ref. 60]; (V) double-stranded RNA; (W) aluminium salts, such as aluminium hydroxides (including oxyhydroxides), aluminium phosphates (including hydroxyphosphates), aluminium sulfate, *etc* [Chapters 8 & 9 in ref. 61]; and (X) other substances that act as immunostimulating agents to enhance the effectiveness of the composition [*e.g.* see Chapter 7 of ref. 45].

The vesicles in the compositions of the invention will be present in 'immunologically effective amounts' i.e. the administration of that amount to an individual, either in a single dose or as part of a series, is effective for treatment or prevention of disease. This amount varies depending upon the health and physical condition of the individual to be treated, age, the taxonomic group of individual to be treated (e.g. non-human primate, primate, etc.), the capacity of the individual's immune system to synthesise antibodies, the degree of protection desired, the formulation of the vaccine, the treating doctor's assessment of the medical situation, and other relevant factors. It is expected that the amount will fall in a relatively broad range that can be determined through routine trials. Dosage treatment may be a single dose schedule or a multiple dose schedule (e.g. including booster doses). The vaccine may be administered in conjunction with other immunoregulatory agents.

Typically, the compositions of the invention are prepared as injectables. Direct delivery of the compositions will generally be parenteral (e.g. by injection, either subcutaneously, intraperitoneally, intravenously or intramuscularly or delivered to the interstitial space of a tissue) or mucosal (e.g. oral or intranasal [62,63]). The compositions can also be administered into a lesion.

Once formulated, the compositions of the invention can be administered directly to the subject. The subjects to be treated can be animals; in particular, human subjects can be treated. The vaccines are particularly useful for vaccinating children and teenagers.

The composition may comprise vesicles from more than one serosubtype of *N.meningitidis* [28]. Similarly, the composition may comprise more than one type of vesicle e.g. both MVs and OMVs

As well as vesicles, the composition of the invention may comprise further antigens. For example, the composition may comprise one or more of the following further antigens:

- antigens from *Helicobacter pylori* such as CagA [64 to 67], VacA [68, 69], NAP [70, 71, 72], HopX [e.g. 73], HopY [e.g. 73] and/or urease.
- a saccharide antigen from *N.meningitidis* serogroup A, C, W135 and/or Y, such as the oligosaccharide disclosed in ref. 74 from serogroup C [see also ref. 75] or the oligosaccharides of ref. 76.
- a saccharide antigen from *Streptococcus pneumoniae* [e.g. 77, 78, 79].
- an antigen from hepatitis A virus, such as inactivated virus [e.g. 80, 81].
- an antigen from hepatitis B virus, such as the surface and/or core antigens [e.g. 81, 82].
- an antigen from *Bordetella pertussis*, such as pertussis holotoxin (PT) and filamentous haemagglutinin (FHA) from *B.pertussis*, optionally also in combination with pertactin and/or agglutinogens 2 and 3 [e.g. refs. 83 & 84].
- a diphtheria antigen, such as a diphtheria toxoid [e.g. chapter 3 of ref. 85] e.g. the CRM<sub>197</sub> mutant [e.g. 86].
- a tetanus antigen, such as a tetanus toxoid [e.g. chapter 4 of ref. 105].
- a saccharide antigen from *Haemophilus influenzae* B [e.g. 75].

- an antigen from hepatitis C virus [e.g. 87].
- an antigen from *N.gonorrhoeae* [e.g. 88, 89, 90, 91].
- an antigen from *Chlamydia pneumoniae* [e.g. refs. 92 to 98].
- an antigen from *Chlamydia trachomatis* [e.g. 99].
- 5 - an antigen from *Porphyromonas gingivalis* [e.g. 100].
- polio antigen(s) [e.g. 101, 102] such as OPV or, preferably, IPV.
- rabies antigen(s) [e.g. 103] such as lyophilised inactivated virus [e.g. 104, RabAvert™].
- measles, mumps and/or rubella antigens [e.g. chapters 9, 10 & 11 of ref. 105].
- influenza antigen(s) [e.g. chapter 19 of ref. 105], such as the haemagglutinin and/or
- 10 neuraminidase surface proteins.
- an antigen from *Moraxella catarrhalis* [e.g. 106].
- an protein antigen from *Streptococcus agalactiae* (group B streptococcus) [e.g. 107, 108].
- a saccharide antigen from *Streptococcus agalactiae* (group B streptococcus).
- an antigen from *Streptococcus pyogenes* (group A streptococcus) [e.g. 108, 109, 110].
- 15 - an antigen from *Staphylococcus aureus* [e.g. 111].
- an antigen from *Bacillus anthracis* [e.g. 112, 113, 114].
- an antigen from a virus in the flaviviridae family (genus flavivirus), such as from yellow fever virus, Japanese encephalitis virus, four serotypes of Dengue viruses, tick-borne encephalitis virus, West Nile virus.
- 20 - a pestivirus antigen, such as from classical porcine fever virus, bovine viral diarrhoea virus, and/or border disease virus.
- a parvovirus antigen e.g. from parvovirus B19.
- a prion protein (e.g. the CJD prion protein)
- an amyloid protein, such as a beta peptide [115]
- 25 - a cancer antigen, such as those listed in Table 1 of ref. 116 or in tables 3 & 4 of ref. 117.

The composition may comprise one or more of these further antigens.

Toxic protein antigens may be detoxified where necessary (e.g. detoxification of pertussis toxin by chemical and/or genetic means [84]).

30 Where a diphtheria antigen is included in the composition it is preferred also to include tetanus antigen and pertussis antigens. Similarly, where a tetanus antigen is included it is preferred also to include diphtheria and pertussis antigens. Similarly, where a pertussis antigen is included it is preferred also to include diphtheria and tetanus antigens. DTP combinations are thus preferred.

Saccharide antigens are preferably in the form of conjugates. Carrier proteins for the conjugates include the *N.meningitidis* outer membrane protein [118], synthetic peptides [119,120], heat shock

35 proteins [121,122], pertussis proteins [123,124], protein D from *H.influenzae* [125], cytokines [126],

lymphokines [126], hormones [126], growth factors [126], toxin A or B from *C.difficile* [127], iron-uptake proteins [128] *etc.* A preferred carrier protein is the CRM197 diphtheria toxoid [129].

Antigens in the composition will typically be present at a concentration of at least 1µg/ml each. In general, the concentration of any given antigen will be sufficient to elicit an immune response against that antigen.

As an alternative to using protein antigens in the composition of the invention, nucleic acid encoding the antigen may be used. Protein components of the compositions of the invention may thus be replaced by nucleic acid (preferably DNA *e.g.* in the form of a plasmid) that encodes the protein.

#### *Methods of treating patients*

The invention provides vesicles of the invention for use as medicaments.

The invention also provides a method of raising an immune response in a patient, comprising administering to a patient a composition of the invention. The immune response is preferably protective against meningococcal disease, and may comprise a humoral immune response and/or a cellular immune response. The patient is preferably a child.

The method may raise a booster response, in a patient that has already been primed against *N.meningitidis*. Subcutaneous and intranasal prime/boost regimes for OMVs are disclosed in ref. 62.

The invention also provides the use of a vesicle of the invention in the manufacture of a medicament for raising an immune response in an patient. The medicament is preferably an immunogenic composition (*e.g.* a vaccine). The medicament is preferably for the prevention and/or treatment of a disease caused by a *Neisseria* (*e.g.* meningitis, septicaemia, gonorrhoea *etc.*).

Methods and uses of the invention may involve administration of vesicles from more than one serosubtype of *N.meningitidis* [*e.g.* ref. 28].

#### *Definitions*

References to a percentage sequence identity between two amino acid sequences means that, when aligned, that percentage of amino acids are the same in comparing the two sequences. This alignment and the percent homology or sequence identity can be determined using software programs known in the art, for example those described in section 7.7.18 of reference 130. A preferred alignment is determined by the Smith-Waterman homology search algorithm using an affine gap search with a gap open penalty of 12 and a gap extension penalty of 2, BLOSUM matrix of 62. The Smith-Waterman homology search algorithm is well known and is disclosed in reference 131.

The term "comprising" means "including" as well as "consisting" *e.g.* a composition "comprising" X may consist exclusively of X or may include something additional *e.g.* X + Y.

The term "about" in relation to a numerical value *x* means, for example,  $x \pm 10\%$ .

The word "substantially" does not exclude "completely" e.g. a composition which is "substantially free" from Y may be completely free from Y. Where necessary, the word "substantially" may be omitted from the definition of the invention.

### BRIEF DESCRIPTION OF DRAWINGS

- 5 Figures 1 and 2 show the presence/absence of (1) protein '287' and (2) protein '741' in bacteria ('TOT') and outer membrane vesicles ('OMV') prepared from strains MC58, H4476 and 394/98 of *N.meningitidis*. The arrow shows the position of '287' in Figure 1 and '741' in Figure 2.

### MODES FOR CARRYING OUT THE INVENTION

#### OMV preparation

- 10 OMVs were prepared either by the prior art 'Norwegian' methods (strains H4476 and 394/98) or by the following process (strain MC58):
- Bacteria from 2-5 plates were harvested into 10 ml of 10mM Tris-HCl buffer (pH 8.0) and heat-killed at 56°C for 45min. The samples were then sonicated on ice (duty cycle 50 for 10 minutes with the tip at 6/7) to disrupt membranes.
  - 15 – Cellular debris was removed by centrifugation at 5000g for 30 minutes at 4°C, or 10000g for 10 minutes.
  - The supernatant was re-centrifuged at 50000g for 75 minutes at 4°C
  - The pellet was resuspended in 7 ml of 2% N-lauroyl sarcosinate (Sarkosyl) in 10 mM Tris-HCl (pH 8.0) for 20 minutes at room temperature to solubilise the cytoplasmic membranes.
  - 20 – The sample was centrifuged at 10000g for 10 minutes to remove particulates and the supernatant was centrifuged at 75000g for 75 minutes at 4°C. The sample was washed in 10mM Tris-HCl (pH 8.0) and centrifuged at 75000g for 75 minutes.
  - The pellet was resuspended in 10 mM Tris-HCl (pH 8.0) or distilled water.
- 25 The bacteria and the OMV preparations were tested by Western blot for the presence of NspA, 287 and 741 (Figures 1 & 2) and results are summarised in the following table:

| Strain | Detergent | NspA     |        | 287      |     | 741      |     |
|--------|-----------|----------|--------|----------|-----|----------|-----|
|        |           | Bacteria | OMV    | Bacteria | OMV | Bacteria | OMV |
| MC58   | –         | +++      | +++    | +++      | +++ | +++      | +++ |
| H44/76 | +         | +++      | – [20] | +++      | –   | +++      | +   |
| 394/98 | +         | +++      | n.d.   | +++      | ++  | +++      | +   |

In contrast to the prior art detergent-based methods, therefore, the absence of detergent results in NspA being retained in OMVs and avoids loss of 287 & 741.

It will be understood that the invention has been described by way of example only and modifications may be made whilst remaining within the scope and spirit of the invention.

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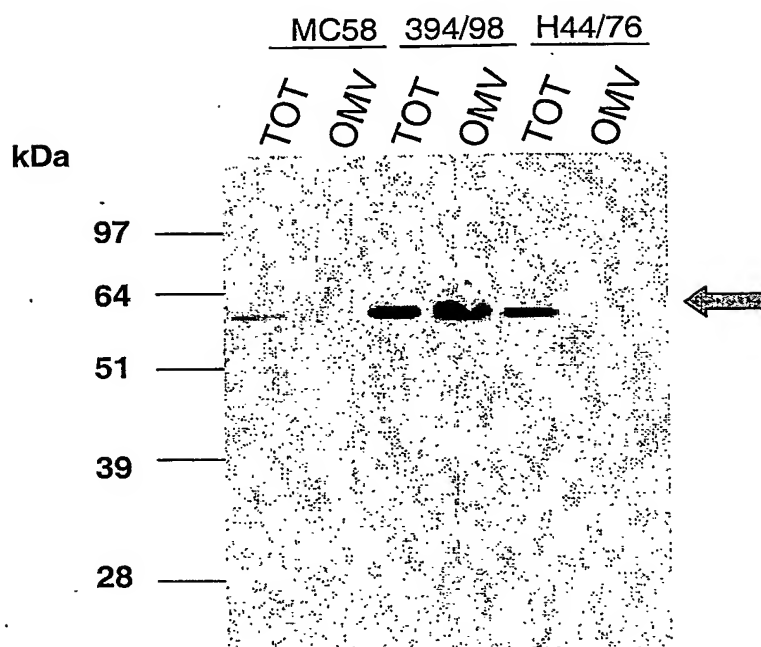
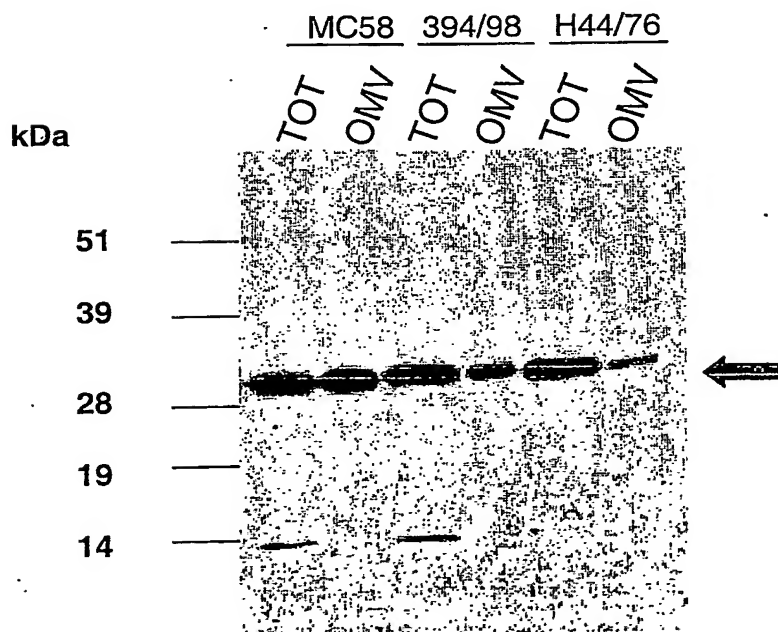
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**FIGURE 1****FIGURE 2**

PCT Application

**IB0304293**

